

Case Report

Acute Renal Failure During a Trial of Spinal Cord Stimulation: Theories as to a Possible Connection

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Objective: This is the first case describing an episode of acute renal failure occurring during a spinal cord stimulation trial.

Clinical Presentation: A 48-year-old male with a history of hypertension and 3 prior failed spine surgeries underwent a trial of spinal cord stimulation for uncontrolled bilateral lower extremity neuropathic pain. Two days after the placement of the percutaneous stimulator lead the patient returned complaining of 3 syncopal episodes. He was found to be hypotensive and in acute renal failure with a creatinine of 8.1 and a BUN of 83.

Intervention: The stimulator lead was immediately removed. The patient was admitted to the intensive care unit and responded promptly to rehydration and placement of a urinary catheter. His renal and urological work-ups revealed no significant abnormalities.

Conclusion: The development of the episode of acute renal failure may have been influenced by the secondary effects of spinal cord stimulation. Since acute renal failure has never been associated with the use of spinal cord stimulation, this singular example does not by itself demonstrate a relationship. However, if future episodes are seen, a link between the 2 events could be drawn. For now, it is not clear if the development of this patient's acute renal failure could, in part, be attributed to the use of the spinal cord stimulator or if it was merely coincidental in nature. We do feel it is useful for the clinician to understand the pathophysiologic changes associated with spinal cord stimulation and to see how, at least in theory, there could be a connection.

Key words: acute renal failure, spinal cord stimulation

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Over the past 30 years, spinal cord stimulation (SCS) has become widely accepted in the medical community as a treatment for refractory, chronic pain conditions. Originally based on Melzack and Wall's "Gate Control Theory of Pain" (1), SCS was first described by Shealy et al (2) back in 1967 to

treat a patient with intractable lower extremity pain. Since then, alternate theories have arisen to explain how SCS works to include activation of second order neurons and inhibitory interneurons at the level of the dorsal horn (3,4), reducing the level of excitatory amino acids via increased gamma amino butyric acid

levels (5), and release of endogenous opioids (6). The indications for SCS have also gradually expanded to pelvic and abdominal pain, malignant pain, brachial plexopathies, central pain, peripheral vascular disease, complex regional pain syndromes, angina, and failed back surgery syndrome (7-10). Whereas initial reports were tempered by high complication and failure rates, the advent of newer and more versatile systems have improved success rates and lowered complication rates (11-13).

Spinal cord stimulation is commonly used to treat nonmalignant, refractory pain that has failed to respond to conservative therapy. Among the various pain conditions that fall into this category, the most common one treated with SCS is failed back surgery syndrome (FBSS) (14). Turner et al (15) recently conducted a systematic review of SCS in patients with FBSS or complex regional pain syndrome). Although the best studies addressed CRPS I patients, there were enough relevant studies to conclude that SCS provides mild to moderate relief of pain in FBSS patients, with leg pain responding better than back pain. The mean complication rate across the 22 studies reviewed was 34.3%, with the most frequently cited ones being the need for stimulator revision (23%) or removal (11%), equipment failure (10%), pain in the region of hardware (5.8%), and superficial infection (4.5%). Less frequently reported complications include aberrant stimulation (16), refractory insomnia (17), atelectasis, pneumonia (18) and increased stimulation in the vicinity of a high-tension electricity substation (19). In this report, we describe a patient with FBSS who developed acute renal failure diagnosed 3 days after a spinal cord stimulator trial. To the best of our knowledge, this complication has never been previously reported. Possible mechanisms for this phenomenon are discussed.

CASE REPORT

A 48-year-old male with a history of hypertension, gastroesophageal reflux, and hepatitis B was referred to the Walter Reed Army Medical Center pain management clinic for evaluation of persistent bilateral lower extremity pain status post-spine surgery. The patient had a complex history of low back and right leg pain dating back to 1979 after a fall aboard a naval ship. Since that time he had undergone extensive treatment ranging from medical management to surgery. His surgeries included a lumbar fusion at L5-S1 with 2 subsequent revisions. After his surgeries he was

left with significant low back and leg pain that was poorly managed with medications and multiple spinal injections.

On initial evaluation, the patient described his low back pain as constant and sharp, with radiation into the right calf and left buttock. He had associated numbness and paresthesias, but denied weakness or bowel/bladder symptoms. His analgesic medications at that time included sustained-release morphine 200 mg po BID, cyclobenzaprine 10 mg po TID, and approximately 90 mg per day of immediate-release morphine for breakthrough pain. His physical examination was remarkable for an intact sensory exam with normal motor strength, an absent left Achille's reflex, and a negative straight leg raise test. Baseline laboratories were not performed, but per patient report a complete blood cell count and full chemistry panel a little more than a year earlier were normal. On the visual analogue pain scale, he rated his pain as an 8/10 at rest, increasing to 10/10 with activity.

After discussing treatment options with the patient and his referring neurosurgeon, the decision was made to proceed with a spinal cord stimulator trial after removal of his bone growth stimulator. In October 2004, the patient had a single lead spinal cord stimulator placed percutaneously (Medtronic Verify® 60 cm lead) in the morning. The patient was on NPO status since the prior evening. During the trial no sedation was administered, although the patient did receive 500 ml of lactated ringer's solution. His vital signs throughout the trial were notable only for a slightly elevated systolic blood pressure in the range of 150 – 160 mm Hg. The skin was entered from a paramedian approach at the L2-3 interspace to enable epidural entry at the L1-2 level. Placement of the lead itself took a single pass and was advanced until the tip reached the middle of the T9 vertebral body. The trial itself took approximately 40 minutes, after which the patient was discharged home with standard post-implant instructions.

Later that evening, the patient experienced a syncope episode while walking to the bathroom, which was presaged by dizziness. His symptoms resolved after he laid down in bed. On POD #1 he continued to feel dizzy and was unable to urinate. On POD #2 he returned to the pain clinic requesting that the stimulator be removed. He could not recall urinating for the previous 2 days. Upon questioning, he stated that he continued to experience lightheadedness

when upright, but that his symptoms resolved with recumbency. After removal of the spinal cord stimulator, the patient was sent to the emergency room for further evaluation. Initial vital signs in the ER were heart rate 88/min, blood pressure 77/34 mmHg, respiratory rate 22/min, temperature 98.6 degrees F, and oxygen saturation 99% on room air. His electrocardiogram was normal. Laboratory data was as follows: Na⁺ 136, K⁺ 3.5, Cl⁻ 95, HCO₃⁻ 24, blood urea nitrogen 83 mg/dl, creatinine 8.1 mg/dl, creatinine clearance of 16ml/min, glucose 102, CPK of 383, MB of 1.7, and troponins < 0.1, with a normal complete blood count. After infusion of 3 liters of Ringer's lactate, his blood pressure increased to 100/56 mmHg, his heart rate declined to 73/min, and he was able to produce a small amount of urine. He was subsequently transferred to the medical ICU where a renal ultrasound showed normal appearing kidneys and a urinalysis was normal. The following day his creatinine had decreased to 3.1mg/dl. By discharge, 2 days after admission, his creatinine decreased to 1.0mg/dl, his blood urea nitrogen was 23mg/dl, and he was voiding spontaneously. Subsequent urological work-up was remarkable only for mild obstruction secondary to prostatic hypertrophy and otherwise normal urodynamics.

Discussion

In this paper, we present a patient with hypertension and failed back surgery syndrome who developed acute renal failure (ARF) during a trial of SCS. While the facts are indisputable, what remains to be answered is what relationship, if any, SCS had on his development of ARF. There are no reports of spinal cord stimulators placed in any location, or for any length of time, precipitating renal failure or azotemia. From this patient's post-recovery laboratory values it is clear that he did not have underlying chronic renal insufficiency. He likely was markedly hypovolemic around the time of the SCS placement. This can be concluded from his electrolytes which show a normal bicarbonate level in the face of a highly elevated BUN which normally is consistent with uremia and a substantial acidosis. This either points to a pre-existing contraction alkalosis or a very rapid and significant renal correction which seems unlikely given his ARF. Laboratory error seems unlikely given the gradual reduction in his creatinine level with fluid resuscitation and the fact that he was oliguric at admission.

Our patient did have several independent factors that put him at risk for the development of ARF. These include some of the medications he was taking at the time of the SCS trial and his history of hypertension. The angiotensin-converting enzyme (ACE) inhibitor lisinopril, the thiazide diuretic hydrochlorothiazide, and high doses of morphine all can adversely effect renal function and volume status (20-22).

While in patients with certain types of nephropathies, ACE inhibitors have been shown to have renoprotective properties (23), paradoxically, in patients with congestive heart failure or severe hypovolemia, the use of an ACE inhibitor may actually precipitate acute renal failure (24). The reported patient had a history of hypertension which was modestly controlled at the time of the procedure. Hypertension itself can lead to a hypovolemic state that can be unmasked in the face of decreased sympathetic input. But it would seem unlikely that the stable use of an ACE inhibitor, a moderate and stable dose of morphine, and mild hypertension alone would lead to the dramatic changes noted in this case report.

Before embarking on a discussion of the effects of SCS on renal function one must identify the etiologic factors that lead to ARF. A recent study on the causes of ARF developing in intensive care unit patients identified the most common contributors to ARF as hypotension, sepsis, nephrotoxic drugs and volume depletion (25).

Any of these factors alone or in combination can lead to ischemic renal failure. The long-term prognosis of ARF is dependent on the speed at which the underlying etiologic factors are corrected. In order to achieve a rapid recovery as witnessed in this case, these factors normally need to be addressed between 2 to 12 hours (26). The patient's post-procedure creatinine level of 8.1mg/dl suggests that this process may have been going on for a more prolonged period of time. However, a more chronic condition might not be expected to resolve so quickly and emphatically. Thus, the key question that arises from this case report is to what extent a precedent condition, volume depletion incurred by the surgery, and the surgery itself each contributed to the ensuing ARF.

SCS is widely acknowledged to affect autonomic equilibrium either via sympathetic inhibition or antidromal activation of sensory fibers (27-31). How exactly these alterations are achieved is not clear. There have been no studies to date evaluating the effects

of lower thoracic SCS on renal blood flow. There are, however, a plethora of clinical and animal studies that show that SCS does indeed produce vasodilation via 2 mechanisms: a decrease in sympathetic outflow and a release of neurohumoral factors that directly lead to relaxation of (micro)vascular smooth muscles.

There is ample evidence that SCS can attenuate sympathetic responses. This idea was first postulated after it was noted that the effects of surgical sympathectomy mimicked the vasodilatory effects of SCS (32). The concept is further supported by animal studies indicating that the vasodilatory effects of SCS are reduced or obliterated by surgical sympathectomy (33). In animal studies evaluating the role cholinergic and adrenergic receptors play in SCS-induced peripheral vasodilatation, Linderoth et al (34) found that the vasodilatory effects of SCS were inhibited by beta and ganglionic blockers, but not muscarinic antagonists. They concluded that the SCS-induced inhibitory effect on peripheral vasoconstriction was maintained via efferent sympathetic activity involving pre- and post-ganglion adrenoreceptors. Increases in sympathetic tone can also result in increased renal arterial pressure (35).

Clinical studies have demonstrated a relative increase in parasympathetic activity affecting visceral organs with spinal cord stimulation. This is demonstrated by the effects of sacral nerve stimulation, a procedure used to decrease bladder hyperactivity and increase bladder capacity. A contributing factor to the beneficial effect SCS exerts on ischemic cardiac muscles has been postulated to be an increase in vagal nerve output (36). Thakkar et al (37) reported the development of a plethora of different gastrointestinal symptoms presumably due to unopposed parasympathetic activity in 2 patients who underwent SCS implantation. The adverse effects that developed were severe enough to require cessation of therapy despite excellent analgesia.

In the normal state, cardiovascular responses to decreased renal blood flow are regulated by a reflex mechanism involving the sympathetic chain at the T9 to L4 levels. This reflex response leads to an elevation in mean arterial pressure and heart rate, which in turn promote increases in renal blood flow and glomerular filtration rate (GFR) (38).

In theory, the decreased sympathetic input induced by SCS could potentially lead to diminished renal blood flow via peripheral shunting, decreased

renal perfusion pressure, and an attenuated cardiovascular response to the ensuing hypotension.

There are several proposed mechanisms for the peripheral vasodilatory effects of spinal cord stimulation. These include antidromic activation of sensory fibers of the vanilloid receptor-1 type (39), increased local release of calcitonin gene related peptide (40), local release of nitric oxide (41), and activation of extracellular signal-regulated kinase (ERK) and protein kinase B (AKT) (42). Despite emerging information regarding these effects, they generally apply to cutaneous vasodilation and hence do not address the effect SCS has on perfusion to other organs and tissues. Exactly how much sympathetic inhibition contributes to hypotension remains unknown. The lone study that addressed the effects SCS has on mean arterial pressures failed to detect a difference in normal volunteers (43). In the present case, we operated on the premise that SCS may have unmasked developing ARF that developed primarily in response to the patient's pre-existing condition and superimposed volume depletion.

Although one cannot rule out coincidence, the timing of the occurrence makes it conceivable that our patient's ARF developed from a confluence of events concluding with SCS. These events include a history of hypertension, the medications he was taking, preoperative hypovolemia exacerbated by NPO status, with further reductions in renal blood flow from the advent of SCS. The postoperative hypotension was probably present on POD #0 as evidenced by the patient's syncopal episode, and persisted for 2 additional days until treatment in the emergency room.

So what can one conclude from this report? We do not believe that hypertension or even pre-existing nephropathy should be a contraindication to SCS. Rather, this case at the very least opens up the discussion on the possible role SCS plays in altering the function of other organs. We believe that SCS should be implemented with caution in patients with renal disease. A more detailed history and physical may reveal developing illnesses or hypovolemia. Some possible steps that may reduce even further the remote risk of renal failure in patients having SCS include obtaining pre- and postoperative electrolytes in patients at risk for kidney disease, maintaining adequate pre- and post-implantation hydration, and alerting patients to seek prompt medical evaluation should they develop any untoward symptoms.

CONCLUSION

In conclusion, we report a case of ARF in a patient with hypertension during a spinal cord stimulator trial. The development of the episode of ARF could have been affected, in part, by the secondary effects of SCS. Since ARF has never been associated with the use of SCS, this singular example does not by itself demonstrate a relationship. However, if future episodes are

seen, a link between the 2 events could be drawn. For now, it is not clear if this case could be attributed to the use of the spinal cord stimulator or if it was coincidental in nature. We do feel it is useful for the clinician to understand the pathophysiologic changes associated with SCS and to see how, at least in theory, there could be a connection.

REFERENCES

- Melzack R, Wall PD. Pain mechanisms: A new theory. *Science* 1965; 150:971-979.
- Shealy CN, Mortimer JT, Reswick JB. Electrical inhibition of pain by stimulation of the dorsal columns: Preliminary clinical report. *Anesth Analg* 1967; 46:489-491.
- Yakhnitsa V, Linderoth B, Meyerson BA. Spinal cord stimulation attenuates dorsal horn neuronal hyperexcitability in a rat model of mononeuropathy. *Pain*. 1999; 79:223-233.
- Dubuisson D. Effect of dorsal-column stimulation on gelatinosa and marginal neurons of cat spinal cord. *J Neurosurg*. 1989; 70:257-265.
- Meyerson BA, Linderoth B. Mechanisms of spinal cord stimulation in neuropathic pain. *Neurol Res* 2000; 22:285-292.
- Tonelli L, Setti T, Falasca A, Martignoni E, Torcia E, Calcaterra FM, Merli GA, Facchinetti F. Investigation on cerebrospinal fluid opioids and neurotransmitters related to spinal cord stimulation. *Appl Neurophysiol* 1988; 51:324-332.
- Meglio M. Spinal cord stimulation in chronic pain management. *Neurosurg Clin N Am* 2004; 15:297-306.
- Ubbink DT, Vermeulen H, Spincemaille GH, Gersbach PA, Berg P, Amann W. Systematic review and meta-analysis of controlled trials assessing spinal cord stimulation for inoperable critical leg ischaemia. *Br J Surg*. 2004; 91:948-955.
- DeJongste MJ. Spinal cord stimulation for ischemic heart disease. *Neurol Res* 2000; 22:293-298.
- Ekre O, Eliasson T, Norrsell H, Wahrborg P, Mannheimer C. Electrical stimulation versus coronary artery bypass surgery in severe angina pectoris. Long-term effects of spinal cord stimulation and coronary artery bypass grafting on quality of life and survival in the ESBY study. *Eur Heart J* 2002; 23:1938-1945.
- Sundaraj SR, Johnstone C, Noore F, Wynn P, Castro M. Spinal cord stimulation: A seven-year audit. *J Clin Neurosci* 2005; 12:264-270.
- Long DM, Erickson DE. Stimulation of the posterior columns of the spinal cord for relief of intractable pain. *Surg Neurol* 1975; 4:134-141.
- Sweet WH. Control of pain by direct electrical stimulation of peripheral nerves. *Clin Neurosurg*. 1976; 23:103-111.
- Kupers RC, Van den Oever R, Van Houdenhove B, Vanmechelen W, Hepp B, Nuttin B, Gybels JM. Spinal cord stimulation in Belgium: A nation-wide survey on the incidence, indications and therapeutic efficacy by the health insurer. *Pain* 1994; 56:211-216.
- Turner JA, Loeser JD, Deyo RA, Sanders SB. Spinal cord stimulation for patients with failed back surgery syndrome or complex regional pain syndrome: A systematic review of effectiveness and complications. *Pain* 2004; 108:137-147.
- Van Buyten JP, Zundert JV, Milbouw G. Treatment of failed back surgery syndrome patients with low back and leg pain: a pilot study of a new dual lead spinal cord stimulation system. *Neuromodulation* 1999; 2:258-256.
- Burton AW, Fukshansky M, Brown J, Hassenbusch SJ. Refractory Insomnia in a Patient with Spinal Cord Stimulator Lead Migration. *Neuromodulation* 2004; 7: 242-245.
- Kumar K, Malik S, Demeria D. Treatment of chronic pain with spinal cord stimulation versus alternative therapies: cost-effectiveness analysis. *Neurosurgery* 2002; 51:106-115.
- Murphy PM, MacSullivan R. Spinal Cord stimulation for refractory angina pectoris: A shocking experience. *Neuromodulation* 2004; 7:246.
- Rothbard RL, Schreiner BF, Yu PN. Hemodynamic and respiratory effects of dezocine, ciramadol, and morphine. *Clin Pharmacol Ther* 1985; 38:84-88.
- Camu F, Van Overberge L, Bullingham R, Lloyd J. Hemodynamic effects of two intravenous doses of ketorolac tromethamine compared with morphine. *Pharmacotherapy* 1990; 10:122S-126S.
- Popio KA, Jackson DH, Ross AM, Schreiner BF, Yu PN. Hemodynamic and respiratory effects of morphine and butorphanol. *Clin Pharmacol Ther* 1978; 23:281-287.
- Andersen NH, Morgensen CE. Dual blockade of the renin angiotensin system in diabetic and nondiabetic kidney disease. *Curr Hypertens Rep* 2004; 6: 369-376.
- Macias-Nunez JF, Fernandez R, Calvo C, Grande J, Herrera J, Bustamante J, Garay R, Robles R, Lopez-Novoa JM. Verapamil reverts acute renal functional impairment induced by angiotensin II converting enzyme inhibitors. *Ren Fail* 2003; 25:727-737.
- Prakash J, Murthy AS, Vohra R, Rajak M, Mathur SK. Acute renal failure in the intensive care unit. *J Assoc Physicians India* 2006; 54:784-788.

26. W. Gordon Walker and Andrew Welton. Oliguria and renal failure. In Harvey AM, Johns RJ, McKusick VA, Owens AH, Ross RS (Eds). *The Principle and Practice of Medicine*. Appleton and Lange. Norwalk, CT, 1988. pp.756-760.
27. Wu M, Linderorth B, Foreman RD. Putative mechanisms behind effects of spinal cord stimulation on vascular diseases: A review of experimental studies. *Auton Neurosci* 2008; 138:9-23.
28. Sagher O, Huang DL. Mechanisms of spinal cord stimulation in ischemia. *Neurosurg Focus* 2006; 21:E2.
29. Tanaka S, Komori N, Barron KW, Chandler MJ, Linderorth B, Foreman RD. Mechanisms of sustained cutaneous vasodilation induced by spinal cord stimulation. *Auton Neurosci* 2004; 114:55-60.
30. Tanaka S, Barron KW, Chandler MJ, Linderorth B, Foreman RD. Role of primary afferents in spinal cord stimulation-induced vasodilation: Characterization of fiber types. *Brain Res* 2003; 959:191-198.
31. Sagher O, Huang DL. Effects of cervical spinal cord stimulation on cerebral blood flow in the rat. *J Neurosurg* 2000; 93(1 Suppl):71-76.
32. Augustinsson LE, Augustinsson CA, Holm CJ, Jivegard L. Epidural electrical stimulation in severe limb ischemia. Pain relief, increased blood flow, and a possible limb-saving effect. *Ann Surg* 1985; 202:104-110.
33. Linderorth B, Fedorcsak I, Meyerson BA. Peripheral vasodilatation after spinal cord stimulation: animal studies of putative effector mechanisms. *Neurosurgery* 1991; 28:187-195.
34. Linderorth B, Herregodts P, Meyerson BA. Sympathetic mediation of peripheral mediation of peripheral vasodilation induced by spinal cord stimulation: animal studies of the role of cholinergic and adrenergic receptor subtypes. *Neurosurgery* 1994; 35:711-719.
35. Gao SA, Ambring A, Lambert G, Karlsson AK. Autonomic control of the heart and renal vascular bed during autonomic dysreflexia in high spinal cord injury. *Clin Auton Res* 2002; 12:457-464.
36. Olgin JE, Takahashi T, Wilson E, Vereckei A, Steinberg H, Zipes DP. Effects of thoracic spinal cord stimulation on cardiac autonomic regulation of the sinus and atrioventricular nodes. *J Cardiovasc Electrophysiol* 2002; 13:475-481.
37. Thakkar N, Connelly NR, Vieira P. Gastrointestinal symptoms secondary to implanted spinal cord stimulators. *Anesth Analg* 2003; 97:547-549.
38. Stella A. The kidney as a sensor: functional evidence. *J Hypertens Suppl*. 1992; 10:S113-S119.
39. Wu M, Komori N, Qin C, Farber JP, Linderorth B, Foreman RD. Sensory fibers containing vanilloid receptor-1 (VR-1) mediate spinal cord stimulation-induced vasodilation. *Brain Res* 2006; 1107:177-184. Epub 2006 Jul 11.
40. Croom JE, Foreman RD, Chandler MJ, Barron KW. Cutaneous vasodilation during dorsal column stimulation is mediated by dorsal roots and CGRP. *Am J Physiol* 1997; 272(2 Pt 2):H950-957.
41. Croom JE, Foreman RD, Chandler MJ, Koss MC, Barron KW. Role of nitric oxide in cutaneous blood flow increases in the rat hindpaw during dorsal column stimulation. *Neurosurgery* 1997; 40:565-570.
42. Wu M, Komori N, Qin C, Farber JP, Linderorth B, Foreman RD. Extracellular signal-regulated kinase (ERK) and protein kinase B (AKT) pathways involved in spinal cord stimulation (SCS)-induced vasodilation. *Brain Res* 2008; Jan 12).
43. Schultz DM, Musley S, Beltrand P, Christensen J, Euler D, Warman E. Acute cardiovascular effects of epidural spinal cord stimulation. *Pain Physician* 2007; 10:677-685.